

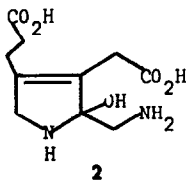
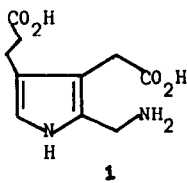
THE CHEMICAL OXIDATION OF PORPHOBILINOGEN

Josefina Awruch and Benjamin Frydman

Facultad de Farmacia y Bioquímica, Universidad de Buenos Aires,
Junín 956, Buenos Aires, Argentina

(Received in USA 19 December 1977; received in UK for publication 29 May 1978)

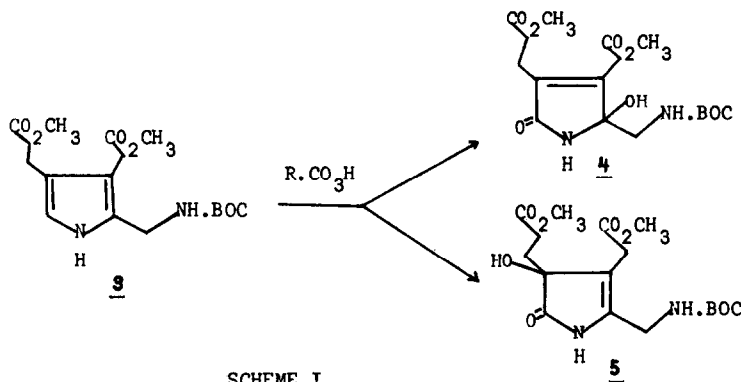
Porphobilinogen (2-aminomethyl-3-carboxymethyl-4-carboxyethylpyrrole) 1 is the universal metabolic precursor of heme, chlorophylls, porphyrins, and the corrin nucleus¹. It is the only known monopyrrole in primary metabolism and has no other biological analogues. It is oxidized



by porphobilinogen oxygenase affording 3-carboxyethyl-4-carboxymethyl-5-aminomethyl-5-hydroxy-3-pyrrolin-2-one (oxoporphobilinogen) 2². We have recently shown that the

amides of δ -keto- α , β -unsaturated acids exist entirely in the cyclic form (as 5-hydroxy-3-pyrrolin-2-ones)³. Hence the structure of 2 is now secured by spectroscopic evidence², by degradation reactions², and by the obtention of synthetic analogues³. Oxoporphobilinogen 2 was found in the urines of porphyric rats excreting urinary porphobilinogen when the rats were treated with pregnenolone to induce hepatic porphobilinogen oxygenase⁴. Since 5-hydroxy-3-pyrrolin-2-ones were detected in urine and blood samples of patients with neurological disorders⁵, and since these disorders are often associated with the various types of porphyrias⁶, it is of interest to obtain 2 in sufficient amounts for pharmacological testing. The chemical oxidation of porphobilinogen 1 appears as the most simple procedure.

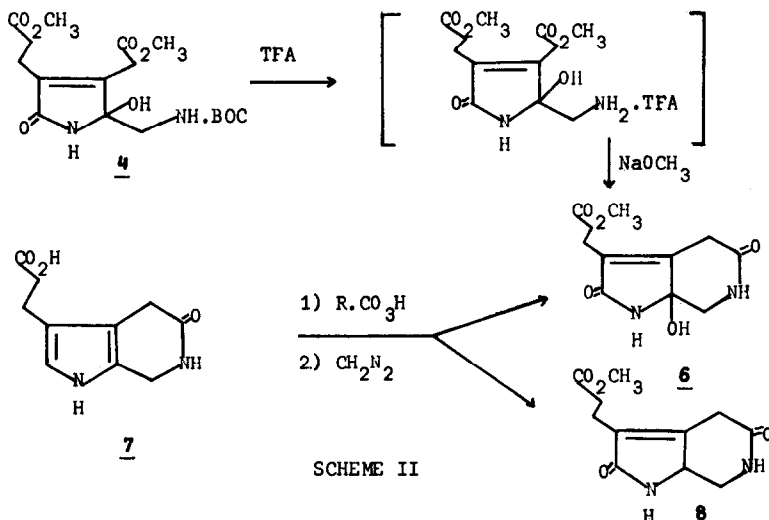
Porphobilinogen 1 can be obtained by a preparative synthesis⁷. We have shown that the peracid oxidation of alkylpyrroles afforded hydroxy-3-pyrrolin-2-one in fair yields⁸. Porphobilinogen 1 was protected prior to the oxidation by treatment with *t*-butyloxycarbonyl azide (50% yield)⁹, followed by esterification of the BOC derivative with diazomethane¹⁰. The BOC dimethyl ester 3 (2g) dissolved in methylene chloride (100ml), was oxidized with two equivalents of *m*-chloroperbenzoic acid at 10° during 2 hs. The solution was washed with a 2% sodium bisulfite solution, then with 5% sodium bicarbonate, dried (Na₂SO₄), evaporated to dryness, and the residue was chromatographed on a tlc silica-gel column packed with 5% methanol in chloroform and was eluted with the same solvent. Two definite oxidation products were obtained (Scheme I).



SCHEME I

The main product (350 mg, 16%) had R_f , 0.50 (tlc on silica-gel; 5% methanol in chloroform) and was the pyrrolinone **4**; ir (Cl_3CH)¹¹: 3450 cm^{-1} (OH); 1740 cm^{-1} (ester CO); 1690 cm^{-1} (amide CO); (nmr; Cl_3CD ; $\delta=0$; TMS); 1.4 (s, 9H, $C(CH_3)_3$), 2.5 (b, 4H, CH_2CH_2); 3.5 (b, 3H, OH, CH_2NH ; reverted to a s 30 min after the addition of D_2O); 3.6 (s, 2H, CH_2CO), 3.7 (s, 6H, OCH_3), 5.2 (t, 1H, CH_2NH); 6.7 (b, 1H, NH); ms (m/e); 386 (M^+ , 18%), 368 ($M-H_2O$, 63%); 369 ($M-OH$, 42%). The second oxidation product had R_f , 0.60 (320 mg, 15%), and was the pyrrolinone **5**; ir (Cl_3CH), 3450 cm^{-1} (broad, bridged OH), 1740 cm^{-1} (ester CO), 1690 cm^{-1} (amide CO); nmr ($CDCl_3$); 1.4 (s, 9H, $C(CH_3)_3$), 2.45, 2.57 (m, 4H, CH_2CH_2), 3.5 (b, 1H, OH; erased after D_2O addition); 3.6 (s, 2H, CH_2CO); 3.64, 3.72 (s, 6H, OCH_3); 4.3 (d, 2H, $J=6\text{HZ}$, CH_2NH ; reverted to a s 30 min after the addition of D_2O); 5.05 (b, 1H, CH_2NH); 6.75 (b, 1H, NH); ms (m/e); 386 (M^+ , 4%), 369 ($M-OH$, 44%).

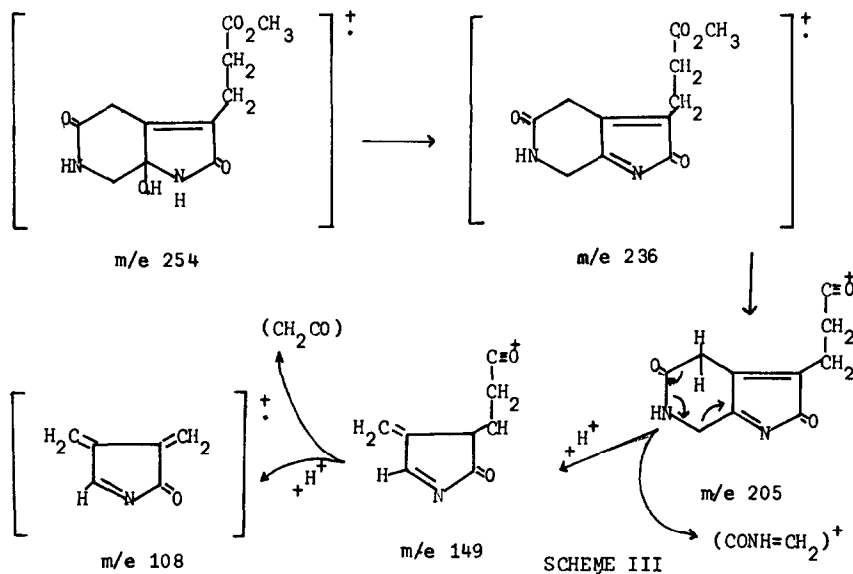
Further confirmation for structure **4** was obtained by converting it into the hydroxylactam **6**, obtained in turn by the peracid oxidation of porphobilinogen lactam **7** (Scheme II).



SCHEME II

The hydroxypyrrolinone **4** was dissolved in trifluoroacetic acid, the solution was kept

at 25° during 20 min; the trifluoroacetic acid was evaporated under nitrogen, and the residue was dissolved in a sodium methoxide solution in methanol and kept at 25° during 24 hs. After evaporation of the solvent, and purification of the residue by chromatography through a tlc silica-gel column using 20% methanol in chloroform, the lactam 6 was obtained as the only product (40% yield). Structure 6 (3-(β -methoxycarbonylmethyl)-2,5-dioxo-7a-hydroxy-2,4,5,6,7,7a-hexahydro-1H-pyrrolo [2,3-c] pyridine) was secured on the basis of its ir (KBr), 1675 cm^{-1} (amide CO, six-membered ring); nmr (Cl_3CD -20% CD_3OD , $\delta=0$, TMS), 2.6 (b, 4H, CH_2CH_2), 3.5 (m, 3H, CH_2CO , OH), 3.7 (s, 3H, OCH_3), 4.0 (b, 2H, CH_2NH); ms (m/e), 254 (M^+ , 10%), 236 (50%), 205 (55%), 207 (30%), 149 (base peak), 108 (40%). The most characteristic fragmentation of the ms spectrum of lactam 6 is depicted in Scheme III and is characteristic for the oxidized lactams of type 6.



Lactam 6 was obtained independently by the chemical oxidation of porphobilinogen lactam 7⁷. The oxidation was carried out in acetic acid using two equivalents of *m*-chloroperbenzoic acid. The oxidation products were esterified with diazomethane and separated by column chromatography on tlc silica-gel using ethyl acetate: methanol, 3:1 as eluant. A substance with R_f , 0.57 was isolated and found to be identical with lactam 6 (31%). The second oxidation product was found to be lactam 8 (47%); ir (KBr), 1750 cm^{-1} (ester CO), 1675 cm^{-1} (amide CO); nmr (TFA, $\delta=0$ for TMS), 2.75 (m, 4H, CH_2CH_2), 3.8 (b, 2H, CH_2CO), 4.0 (s, 3H, OCH_3), 3.8-4.3 (m, 3H, NHCH_2CH); ms (m/e), 238 (M^+ , 90%), 209 (238- CH_2NH , 45%), 149 (238- OCH_3 - CONHCH_2 , base peak), 108 (70%), (see SCHEME III) No 1,2-oxidation product was isolated in the peracid oxidation of 7 or of analogous lactams.

The butyloxycarbonyl group of 4 was cleaved by treatment with trifluoroacetic acid at

room temperature during 20 min. After evaporation of the acid the residue was dissolved in a 2N sodium hydroxide solution which was kept at 20° during 72 hs. The solution was adjusted to pH 5 with acetic acid and filtered through a Dowex-1X4 resin column. Elution with 0.8M acetic acid, and evaporation of the eluates afforded oxoporphobilinogen 2 (42% yield). Its was identical (tlc chromatography) with the product obtained in the enzymatic oxidation of porphobilinogen 1²; ir (Nujol); 3900 cm⁻¹ (OH), 3200 cm⁻¹ (NH); 1680 (CO pyrrolinone); nmr (D₂O, δ=0 for sodium DSS); 2.06 (b, 4H, CH₂CH₂), 2.54 (s, 2H, CH₂COOH); 3.39 (t, 2H, CH₂NH₂), 3.25 (b, 1H, OH); 8.4 (b, 1H, NH); ms (direct inlet, 60°), 258 (M⁺, 40%). The analogous treatment of 5 with trifluoroacetic acid followed by sodium hydroxide did not afford an isolable product. Oxoporphobilinogen 2 was also obtained by saponification of the lactam 6 following a similar procedure to the described for 4.

REFERENCES

1. B. Burnham in Metabolic Pathways, III, (DM. Greenberg Ed. 3rd ed, Acad. Press), p. 403 (1969).
2. R.B. Frydman, M.L. Tomaro, A. Wanschelbaum, E. Andersen, J. Awruch, and B. Frydman, Biochemistry, 12, 5253 (1973).
3. J. Awruch and B. Frydman, Tetrahedron Lett., 4121 (1976).
4. R.B. Frydman, M.L. Tomaro, B. Frydman, and A. Wanschelbaum, FEBS Lett., 51 (1), 206 (1975).
5. D. Irvine, Int. Rev. of Neurobiology, 16, 374 (1974).
6. D.P. Tschudy in Duncan's Diseases of Metabolism (P.K. Bondy and L.E. Rosenberg Ed. W.B. Saunders Comp), p. 775 (1974).
7. B. Frydman, S. Reil, M.E. Despuj, and H. Rapoport, J. Am. Chem. Soc., 91, 2738 (1969).
8. J. Awruch, and B. Frydman, Tetrahedron Lett., 2611 (1973).
9. B. Frydman, S. Reil, A. Valasinas, R.B. Frydman, and H. Rapoport, J. Am. Chem. Soc., 93, 2738 (1971)
10. A.R. Battersby, M. Ihara, E. McDonald, J. Saunders, and R.J. Wells, J. Chem. Soc. Perkin I, (3) 283 (1976).
11. All nuclear magnetic resonance spectra were run on a Perkin-Elmer R-12 instrument; infrared spectra were recorded on a Perkin-Elmer 21 instrument.

Acknowledgement. This work was generously supported by grants from the National Institutes of Health (GM-11973) and the Consejo Nacional de Investigaciones (Argentina).